

# The calcification process and measurement techniques

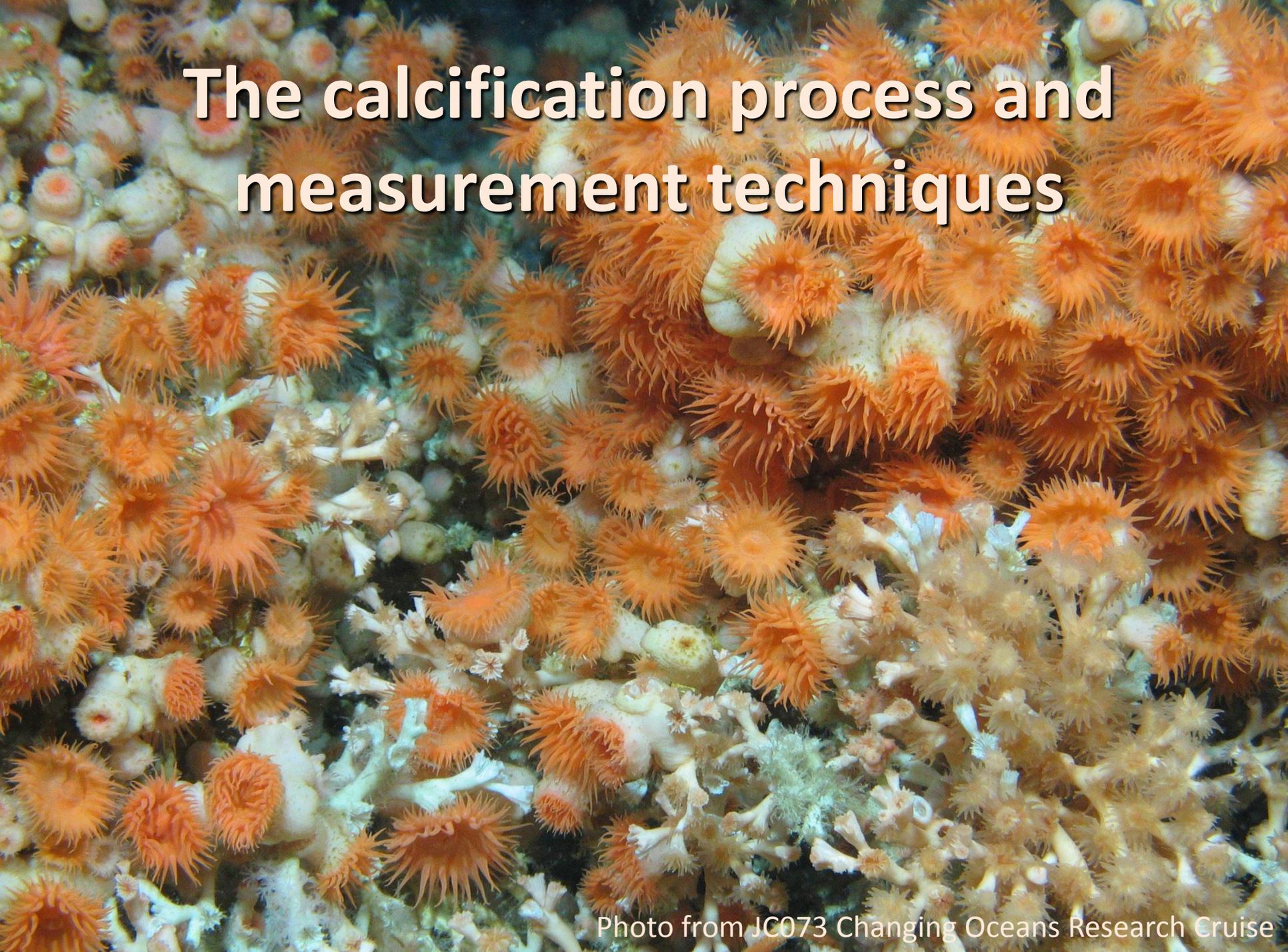
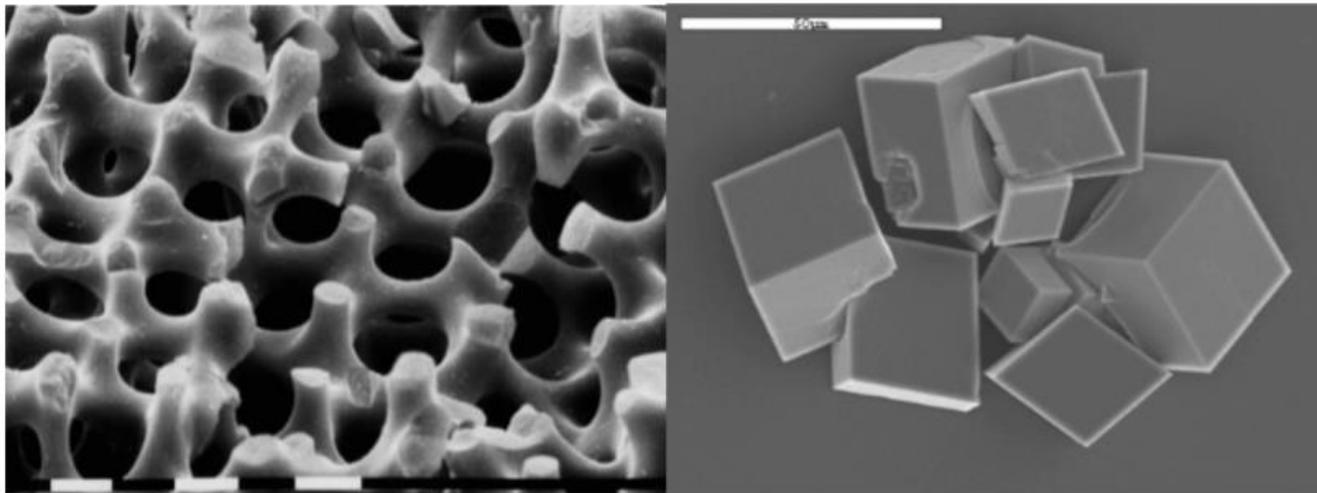
A close-up photograph of a large colony of sea anemones. The anemones are densely packed, with many showing bright orange tentacles and others appearing more pale or white. They are growing on a light-colored, porous rock surface. The background is dark, suggesting an underwater environment.

Photo from JC073 Changing Oceans Research Cruise

# What is calcification?

- The accumulation of calcium salts into body tissue, such as bones, shells, and carapaces.
- A biologically-mediated process
- In marine calcifiers, calcification predominantly results in calcium carbonate structures that are made of either calcite, aragonite or high-Mg calcite.



**Figure 1.** Comparison of calcite single crystals: (*left*) stereom of echinoderm and (*right*) synthetically produced rhombohedral forms.

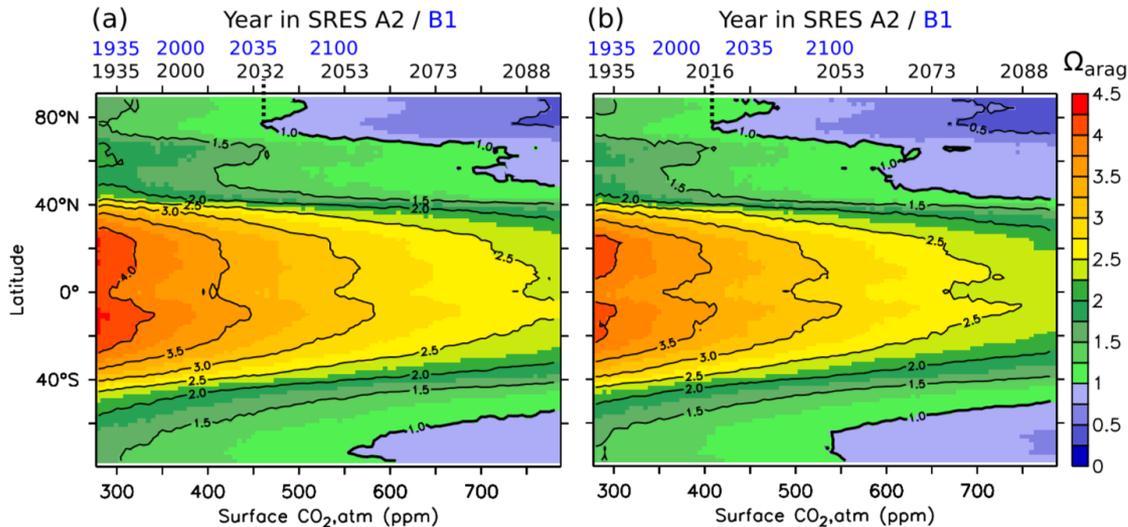
# What is calcification?



**Saturation State** – degree to which seawater is saturated (or not) with relevant ions; provides a measure of the thermodynamic potential for the mineral to form or to dissolve

$$\Omega = \frac{[\text{Ca}^{2+}][\text{CO}_3^{2-}]}{K_{sp}}$$

$\Omega > 1$  Supersaturated with respect to  $\text{CaCO}_3$   
 $\Omega < 1$  Undersaturated with respect to  $\text{CaCO}_3$  (dissolution)



# Who calcifies, and how?

Major invertebrate calcifying groups:

- Molluscs
- Cnidarians
- Echinoderms
- Crustaceans

Other organism types:

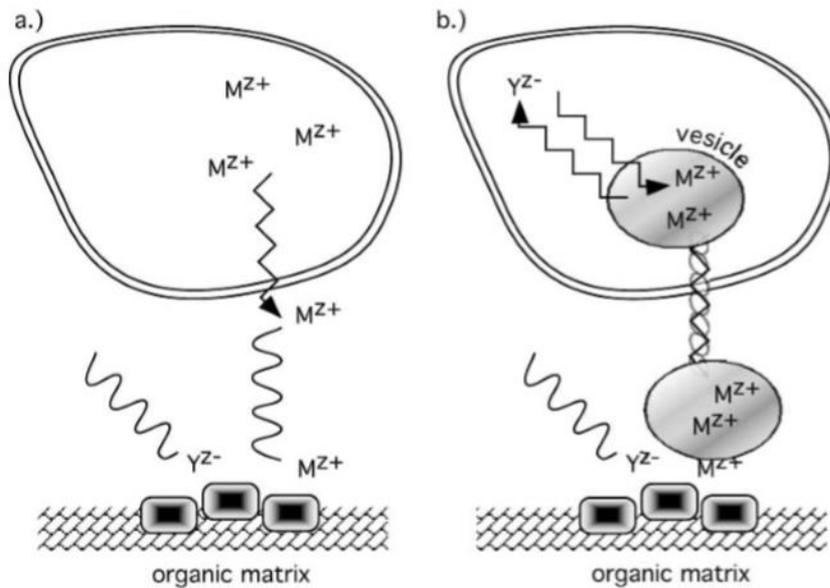
- Formaminifera
- Phytoplankton: Haptophytes (coccolithophores)
- Algae: Rhodophytes (coralline algae)

In most biological systems, the **site of mineral deposition is isolated** from the environment, the extent of isolation is variable.

**Biologically induced mineralisation** – organism uses cellular activities to direct the nucleation, growth, morphology, and final location of the mineral that is deposited. Several types, but most  $\text{CaCO}_3$  forming marine organisms either use an **extracellular** biologically-controlled process or an **intracellular** strategy.

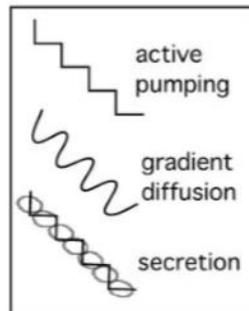
# Who calcifies, and how?

Extracellular biologically-controlled process e.g. Molluscs, Corals,



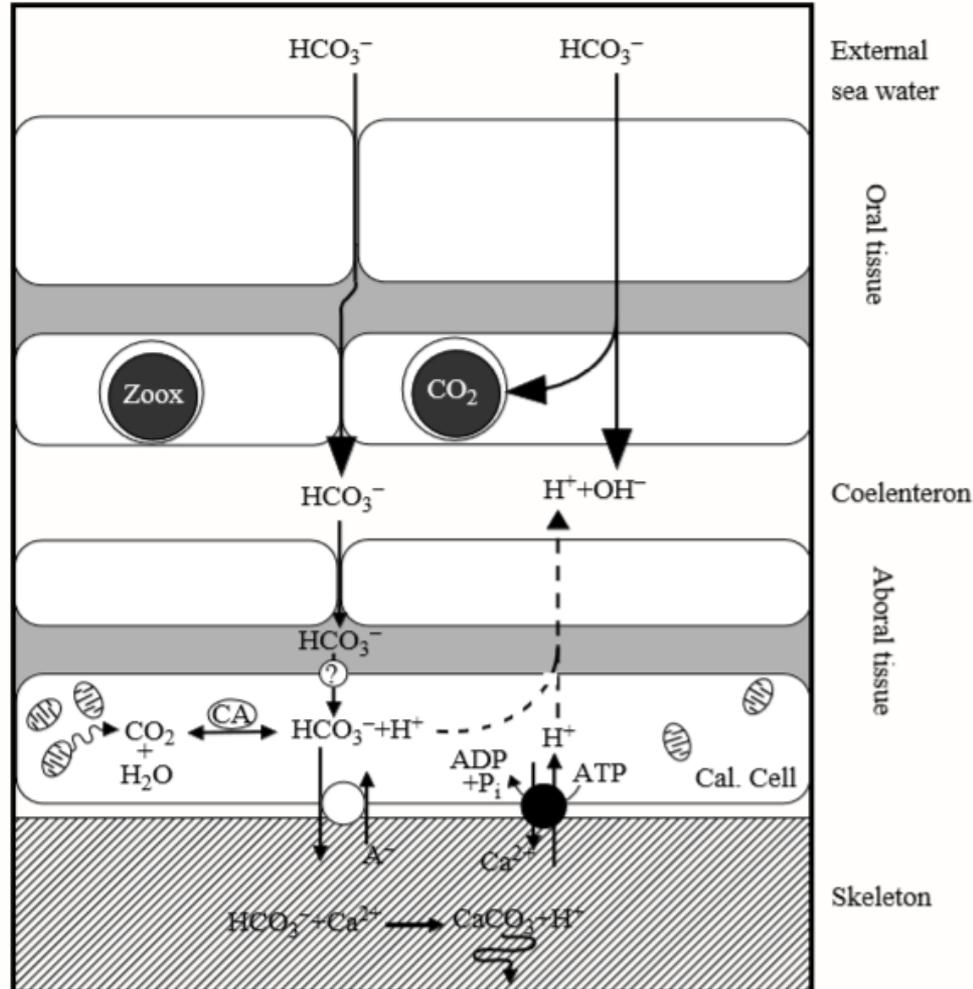
- Basic form of calcification
- Organic matrix important for defining structure
- Ions can be actively pumped out of the cell *or* pumped into a vesicle within the cell which is then secreted outside.

**Figure 5.** Illustrations of biologically controlled extracellular mineralization showing that this process is distinguished by nucleation outside of the cell. a.) Cations are pumped across the cell membrane and move by passive diffusion through extracellular fluids to the site of mineralization. b.) Cations are concentrated intracellularly as aqueous ions into a vesicle that is subsequently secreted. Compartment breakdown at site of mineralization releases cations for biomineral formation.



# Who calcifies, and how?

e.g. Corals

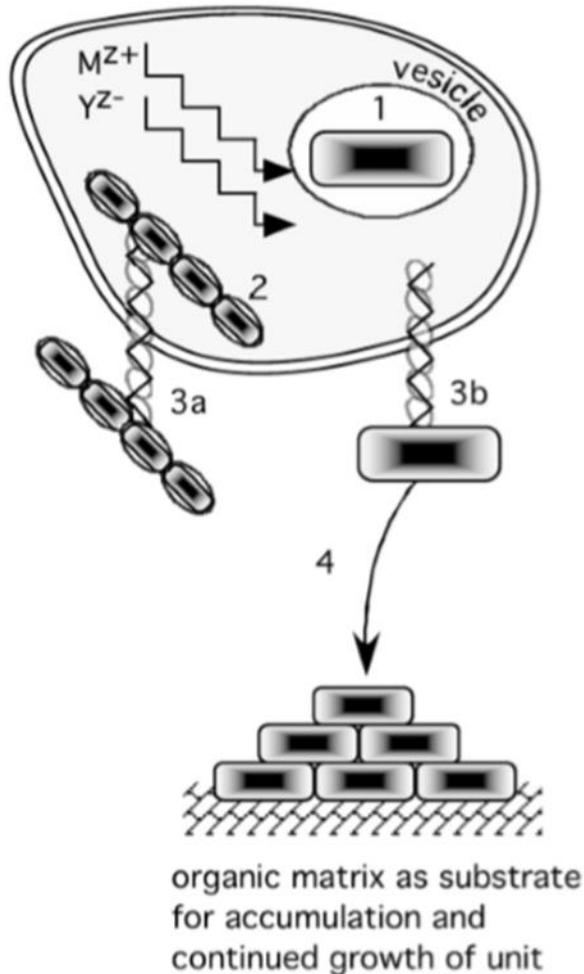


- Model of dissolved inorganic carbon (DIC) absorption for coral calcification and photosynthesis.
- Extracellular space has controlled pH environment
- Anion exchange pumps are utilised for control

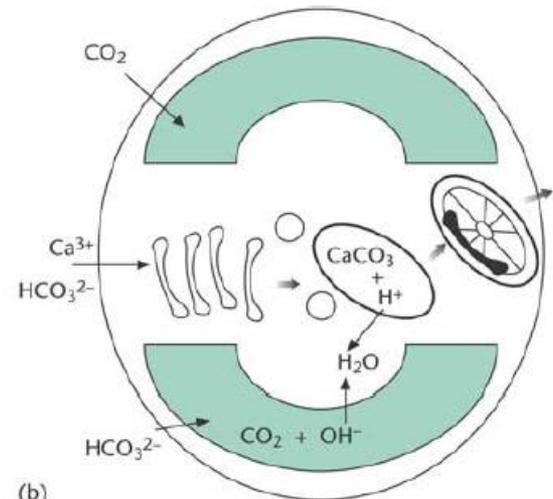
# Who calcifies, and how?

**Intracellular strategy.** E.g. Echinoderms (urchins), coccolithophores...

- Can form huge mineralised products within a vesicle that is the product of many cells fusing their membranes.
- Mineral is exposed to the environment only when the membrane is degraded.



(a)



(b)

# Who calcifies, and how?



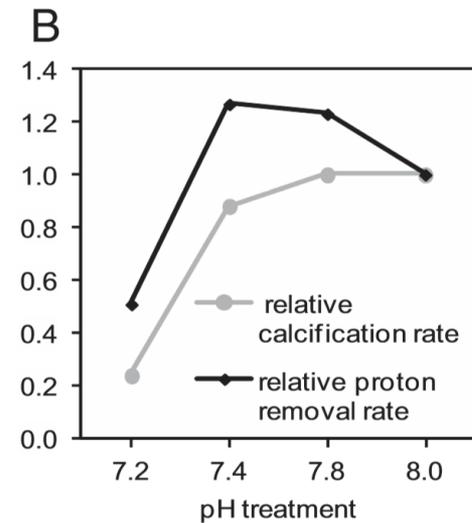
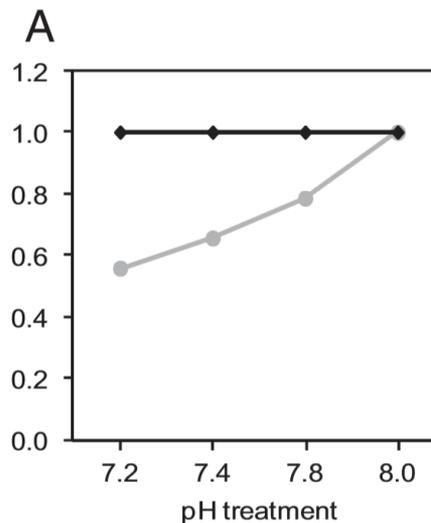
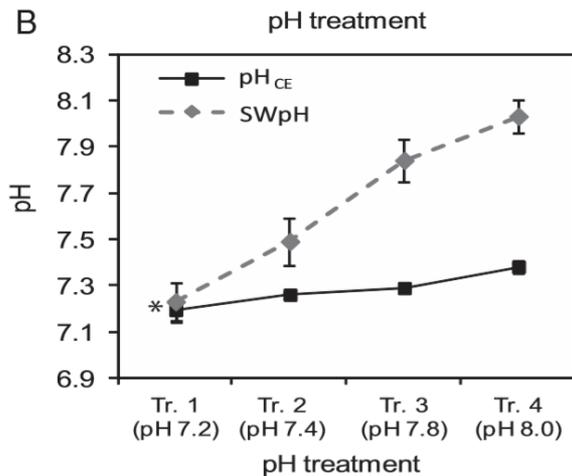
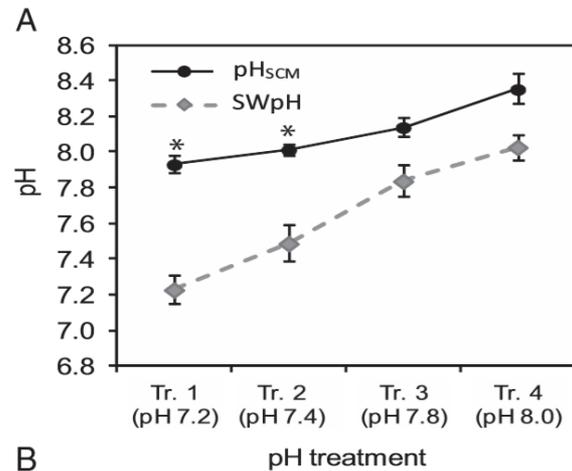
Pane & Barry 2007; Photo MBARI (2006)

- Crustaceans have complex moult cycles
- Able to reabsorb minerals from 'old' shell to incorporate into 'new' shell
- High organic component, as well as chitin
- Organic matrix important for structuring mineral formation
- Different parts of crustaceans (e.g. claws, carapace, legs) have different mineral content which determines 'hardness' and strength

# Why should ocean acidification impact calcification?

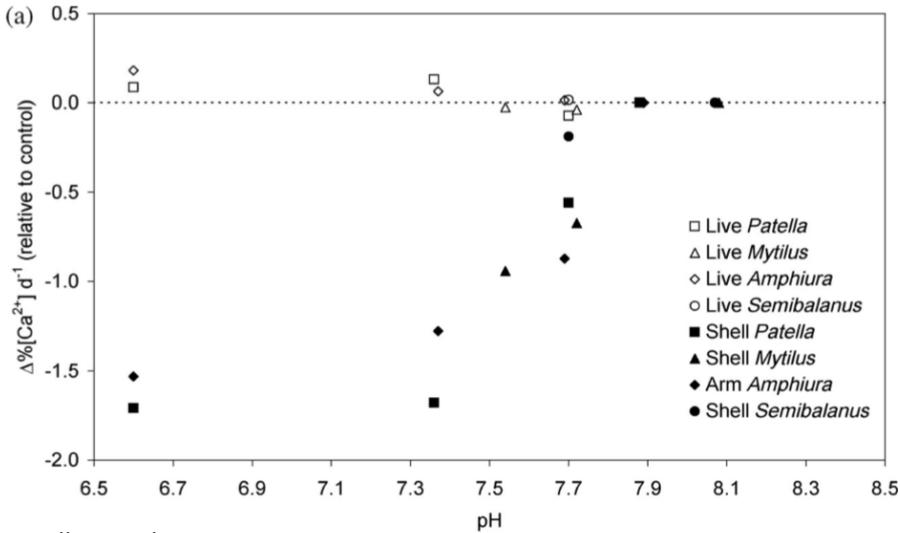
**1. Direct shifts in acid-base balance** (pH, ionic composition) of intracellular fluids that compromise calcification process

e.g. Corals

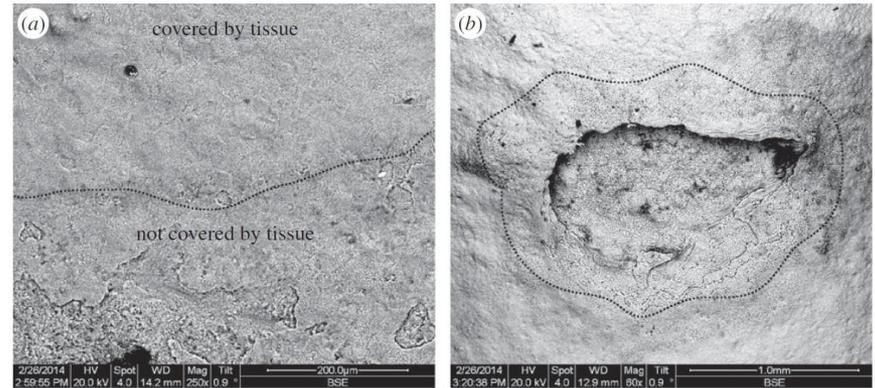


# Why should ocean acidification impact calcification?

**2. Enhanced dissolution** in undersaturated conditions  
 e.g. dissolution of “dead” structures compared to “live”

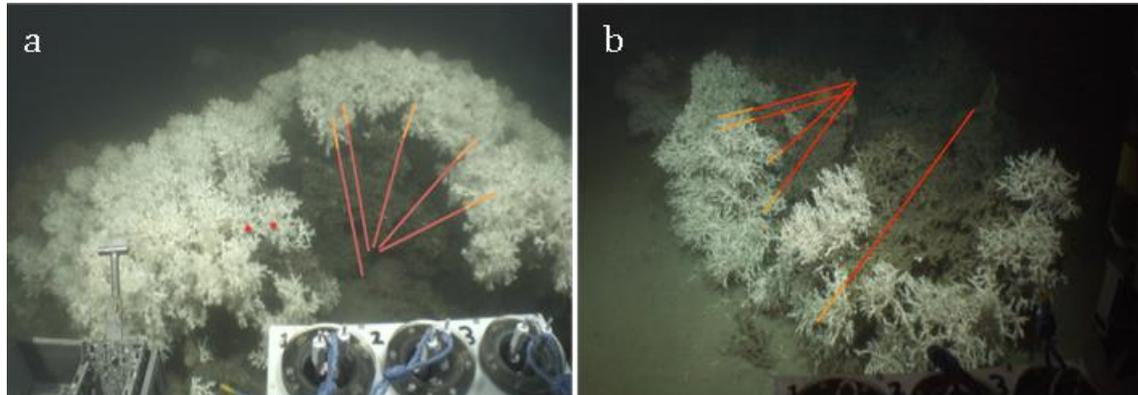


Findlay et al. 2011



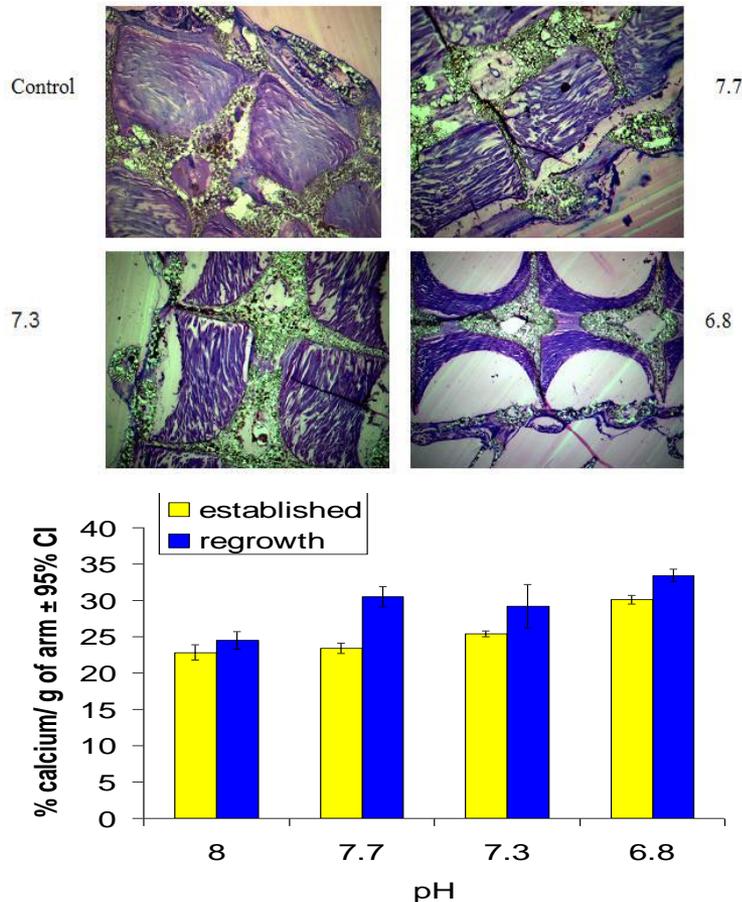
Hennige et al. 2015

Vad et al. in review

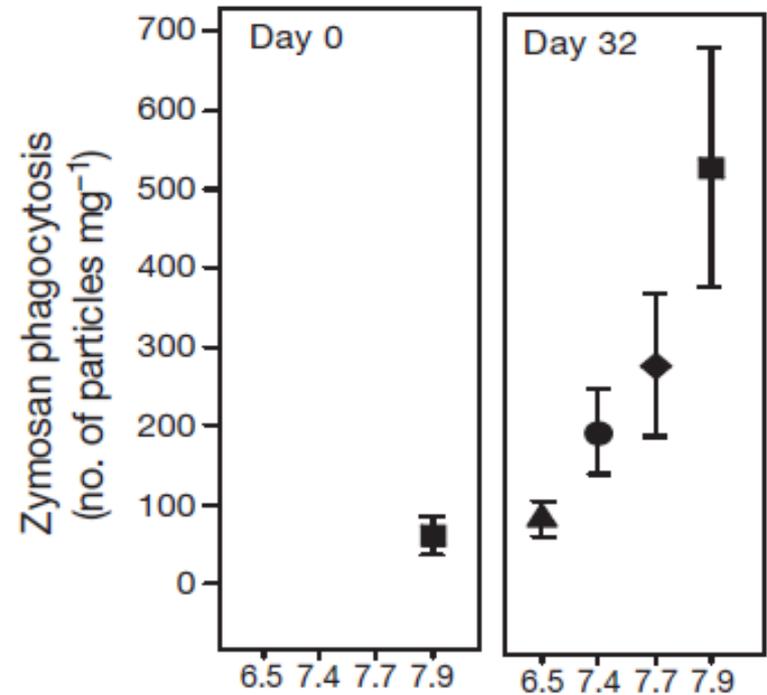


# Why should ocean acidification impact calcification?

**3. Additional energy requirements** needed for maintaining and producing calcium carbonate material in unfavourable conditions  
 e.g. trade-offs between physiological process... brittlestars, mussels, many others...



Wood et al. 2008



Bibby et al. 2008

# Some definitions

- ***Gross calcification***  $\text{CaCO}_3$  precipitated by an organism or community
- ***Net calcification***  $\text{CaCO}_3$  precipitated by an organism or community minus dissolution of  $\text{CaCO}_3$  from the organism or community.
- ***Potential calcification*** Gross calcification, assuming that the organisms considered cover 100% of the area
- ***Net accumulation*** Amount of  $\text{CaCO}_3$  precipitated locally plus the amount of material imported minus dissolution and export

# Summary of techniques

- Geological approach
- Sedimentological approach
- Alkalinity Anomaly Technique
- pH-O<sub>2</sub>
- Change in calcium concentration
- Radioisotopes (<sup>45</sup>Ca, <sup>14</sup>C, <sup>3</sup>H-tetracycline)
- Changes in particulate calcium content
- X-ray analysis
- Buoyant weight
- “Biological” approach
- Changes in Particulate Inorganic Carbon content
- Molecular tools

# Geological

$\text{CaCO}_3$  accumulates in sediment over long time periods giving an indication of rates of calcification.

**Net accumulation of  $\text{CaCO}_3$**  is calculated by the thickness of the layer multiplied by the density, divided by the time increment (measured by radiocarbon dating)

**Level:** Community

**Timescale:** 1000-20000 years

**Examples:** Chave et al. (1972)

**Pros:** Provides integrated, long-term estimates

**Cons:** Numerous uncertainties and assumptions. Highly constrained by sea level



# Sedimentological

Calcified organisms accumulate within sediments. **Net calcification (?)** is measured using the percentage weight contribution in sedimentary skeletal components

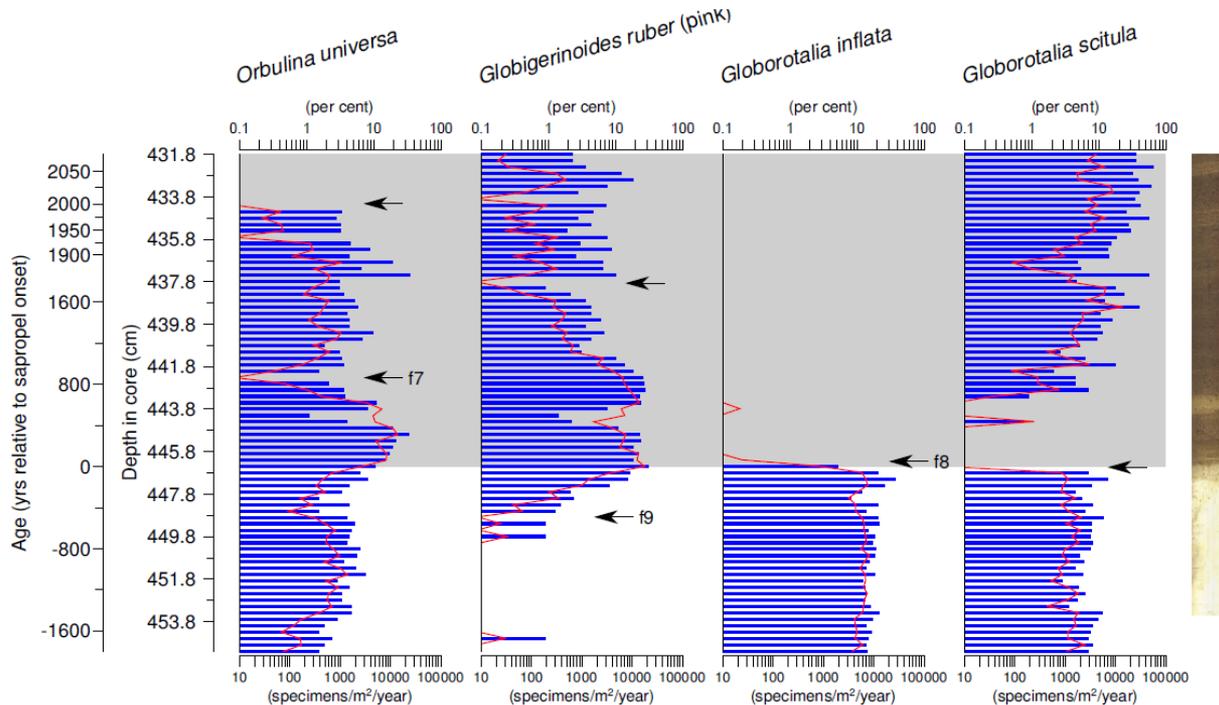
**Level:** Community

**Timescale:** Months

**Examples:** Langer et al. (1997), Wienkauf et al. 2013

**Pros:** Only needs sediment samples.

**Cons:** It is not clear what this approach measures, it does not account for advection terms



# Alkalinity Anomaly Technique

Alkalinity is lowered by two equivalents for each mole of  $\text{CaCO}_3$  precipitated.

**Net calcification** is calculated by measuring the TA before and after an incubation period, and the  $\Delta\text{TA}$  is scaled to  $\Delta\text{CaCO}_3$  (i.e. calcification =  $0.5 \times \Delta\text{TA}$ )

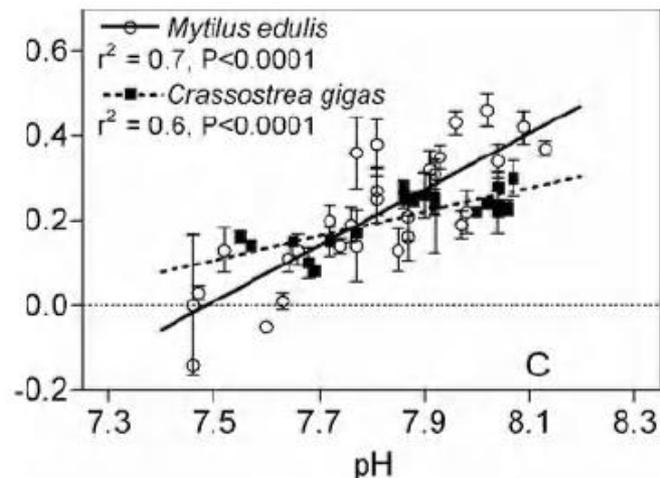
**Level:** Organisms and communities

**Timescale:** Hours to weeks

**Examples:** Smith & Key (1975), Gazeau et al. (2007), Martin et al. (2013), Inoue et al. (2013)

**Pros:** Very precise (1 SD =  $3 \mu\text{mol/kg}$  or about 0.2%)

**Cons:** Needs discrete samples (but see Watanabe et al., 2004). A correction for changes in nutrients may be needed. Need to enclose or know residence time.



# pH-O<sub>2</sub>

Relationships exist between  $\Delta O_2$  and  $\Delta DIC_{org}$ , the metabolic quotients.

**Net calcification** can be measured by estimating net community production and respiration from changes in the concentration of dissolved O<sub>2</sub>.  $\Delta DIC_{calc}$  is then calculated by subtracting  $\Delta DIC_{org}$  from the upstream DIC value.  $\Delta DIC_{calc}$  can be converted to  $\Delta TA$  and consequently calcification.

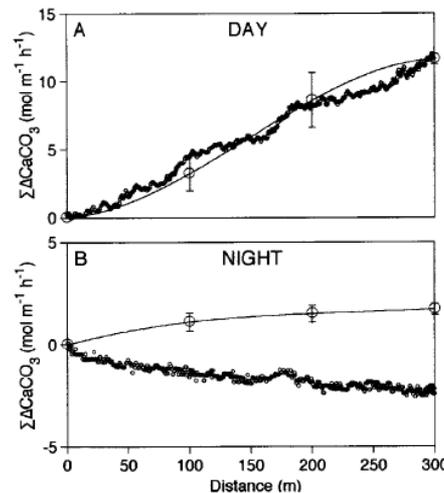
**Level:** Organisms and communities

**Timescale:** Hours

**Examples:** Chisholm & Barnes (1998), Barnes (1983)

**Pros:** It does not require TA monitor (which is timely)

**Cons:** Needs DIC (hence TA) upstream. Assumes metabolic quotients



Chisholm & Barnes 1998

# Calcium concentration

Calcium concentration can directly be measured within internal fluids of organisms. **Net calcification** can be estimated from calcium removal measured using chemical titrations or sensors

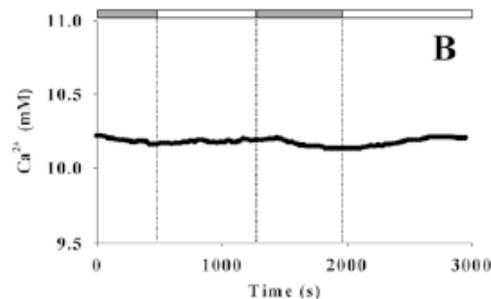
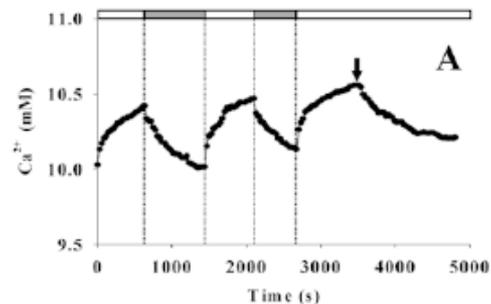
**Level:** Organisms and communities

**Timescale:** Minutes to weeks

**Examples:** Chisholm & Gattuso (1991), Al-Horani et al. (2003)

**Pros:** Direct measurement of calcium uptake; no major assumptions

**Cons:** Low detection limit, high background concentration (10 mmol/l)



# Radio isotopes

Calcium is taken up into the organisms skeletal components, the calcium uptake can be measured using radiolabelled elements ( $^{45}\text{Ca}$ ,  $^{14}\text{C}$  and  $^3\text{H}$ ) to estimate **net calcification**

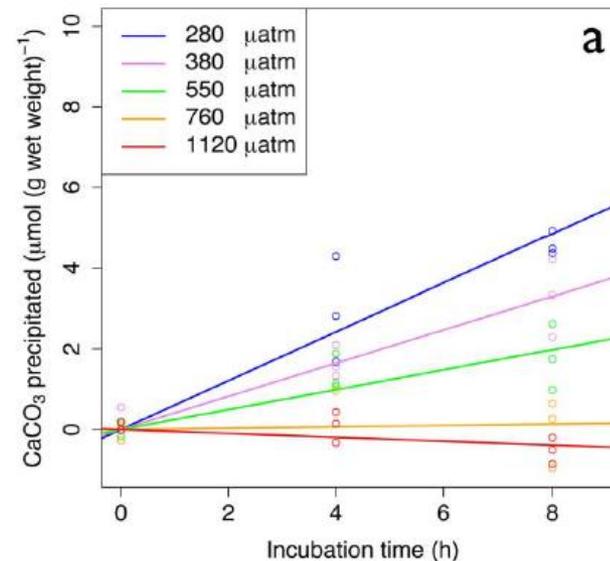
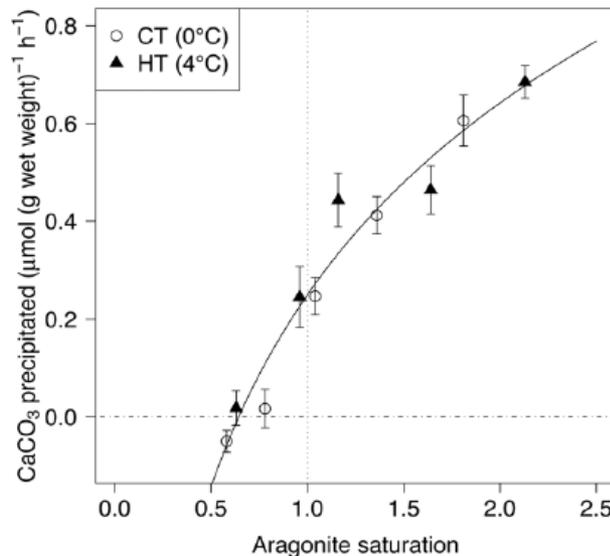
**Level:** Organisms

**Timescale:** Minutes to hours

**Examples:** Fabry et al. (1989), Comeau et al. 2010

**Pros:** Extremely sensitive, Short-term incubations

**Cons:** Destructive, Non-biological adsorption, Use of radioisotopes restricted



# Changes in particulate calcium

Calcium is taken up into the organisms skeletal components, the calcium concentration can be measured by flame atomic absorption spectroscopy to give an estimate of **net calcification**.

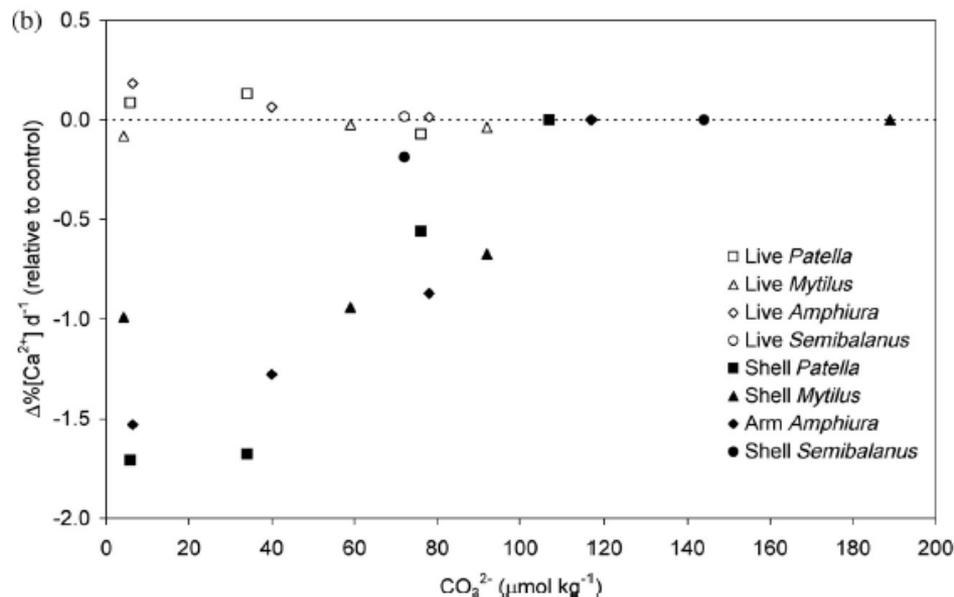
**Level:** Organisms

**Timescale:** Hours to days

**Examples:** (Stoll et al., 2002); (Findlay et al. 2011)

**Pros:** Precision is adequate when growth rates are high (cultures)

**Cons:** Analytical care Instrumentation



# X-rays

X-rays (and Computerised tomography (CT) scanning) measure the density and mass of skeleton, providing a direct measure of **net calcification**, particularly through time (using long-lived coral structures).

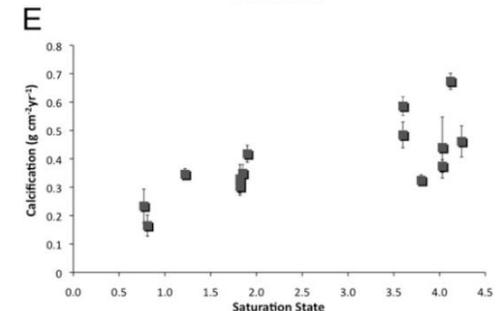
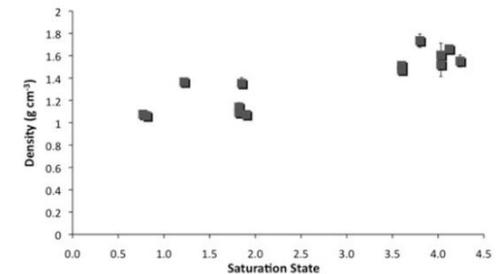
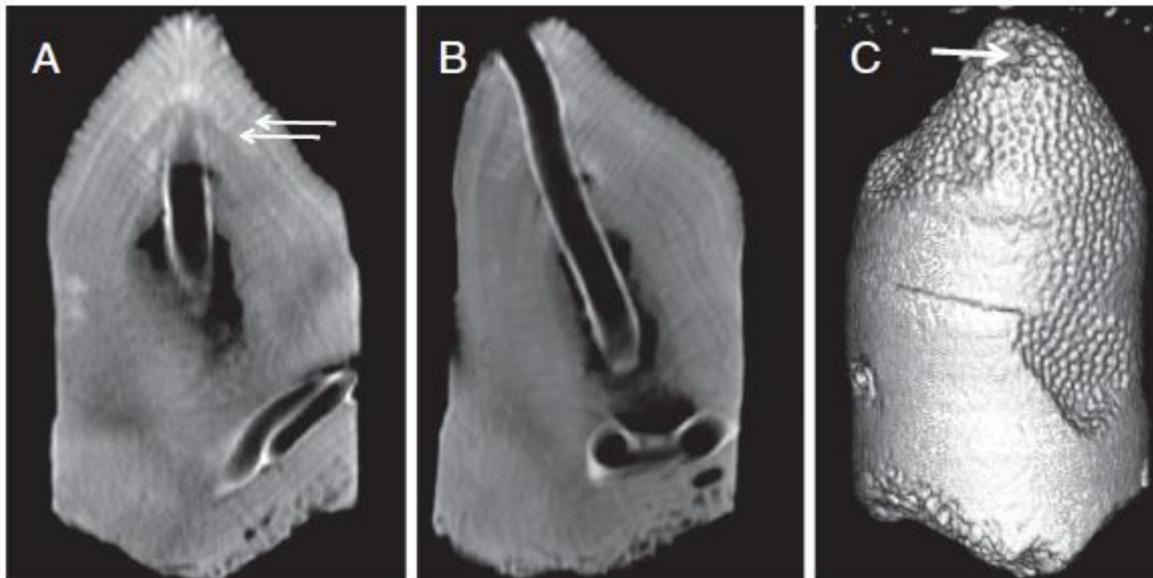
**Level:** Organisms

**Timescale:** days, months, to 100s years

**Examples:** Lough & Barnes (2000), Crook et al. (2013)

**Pros:** Enables retrospective analysis, provides an assessment of erosion

**Cons:** Requires substantial equipment & instrumentation



# Buoyant weight

Increases in mass of an organisms skeleton directly correspond to increases in **net calcification**.

**Level:** Organisms

**Timescale:** Sub-daily to months/years

**Examples:** Dodge et al. 1984, Jokiel et al. 2008

**Pros:** Quite sensitive, Not destructive, No incubation required

**Cons:** Serious problem of normalization for comparative analysis



Dodge et al. 1984

# Biological approaches

Growth measurements or turnover rates (for populations) are associated with an increase in mass of calcified structure and can be used to estimate **net calcification**. Techniques can include using fluorescent dyes (e.g. calcein staining) to observe specific growth areas.

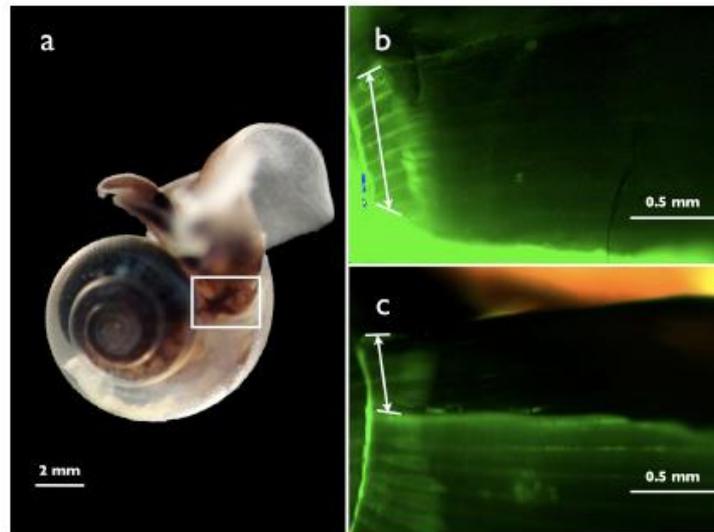
**Level:** Organisms

**Timescale:** Days, months to years

**Examples:** Fabry (1990), Smith (1972), Migné et al. (1998), Comeau et al. (2009)

**Pros:** Simple, individual level

**Cons:** Short term growth not always significant, lots of variability



# Changes in PIC

Changes in the content of the particulate carbon content of an organism reflect its accumulation or loss of carbon and provide an estimate of **net calcification**.

Total particulate carbon (TPC) and particulate organic carbon (POC) are measured (CHN analyzer, mass spectrophotometry).  $PIC = TPC - POC$ .

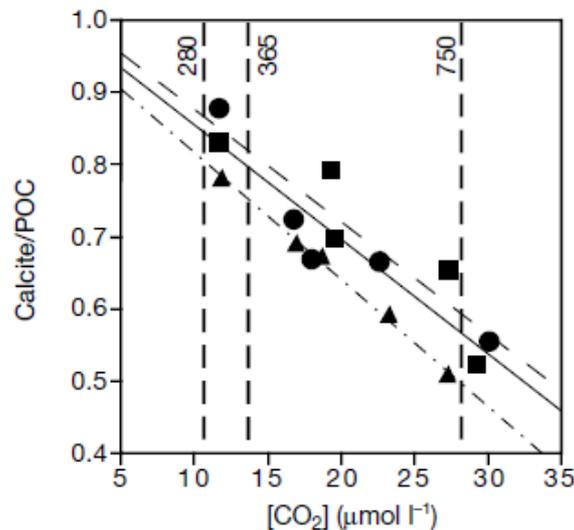
**Level:** Organisms

**Timescale:** Hours to days

**Examples:** Riebesell et al. (2000), Sciandra et al. (2003)

**Pros:** Adequate with cultures and field samples (?)

**Cons:** Instrumentation, Not amenable to automation



# Molecular

Genetics controls the calcification process, by measuring the activity of genes involved in the calcification process (measure mRNA) gives an idea of the **gross calcification (?)**

**Level:** Organisms, perhaps communities?

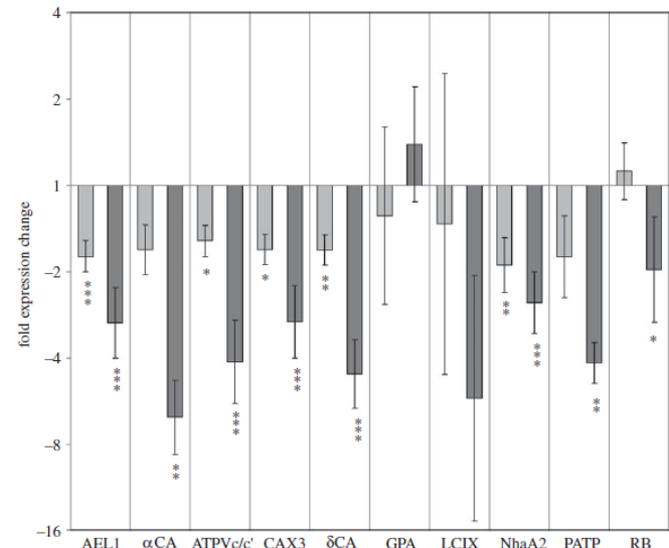
**Timescale:** Hours (to days?)

**Examples:** Lohbeck et al. 2014

**Pros:** High sampling rate because no incubation required

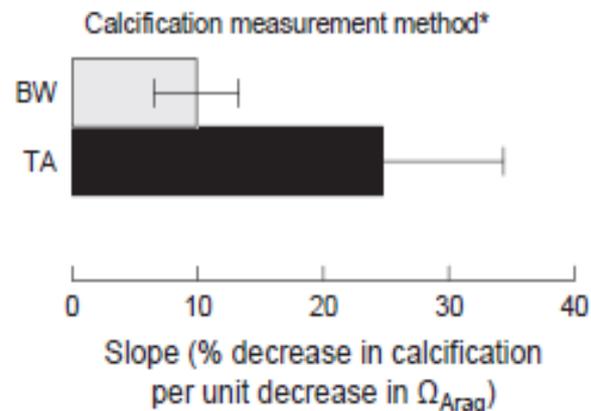
**Cons:** Post-translational regulation, Poor precision (semi-quantitative), Reliance on instrumentation (quantitative real-time PCR), not clearly related to actual production of calcium carbonate skeleton.

Gene name	Full name	Protein ID/GenBank accession number	Putative function	Primer name	Primer sequence 5'-3'	Amplicon size	Reference
EFB1	Elongation Factor1	402437	endogenous reference gene	EFB1_F	GCT GGA AGA AGG ACT TTG TTG	101	Madkinder et al. 2011
				EFB1_R	TCC ACC AGT CCA TGT TCT TC		
Actin	Actin	564388.1*, 564389.1*, 564392.1*, 564393.1*, 564390.1*, 564389.1*	endogenous reference gene	Actin_F	GAC CGA CTG GAT GGT CAA G	96	Madkinder et al. 2011
				Actin_R	GCC AGC TTC TCC TTG ATG TC		
αTUB	α Tubulin	multiple copy	endogenous reference gene	αTUB_F	GCA TCG CCG AGA TCT ACT C	84	Bach et al. 2013
				αTUB_R	TCC CCG ACG TAC CAG TG		
RB	Rubisco	D43845.1	Gene coding for large subunit of RUBISCO	RB_F	CAA TGG GTG ACC CAG ATG GTA	100	Ruhn et al. 2010
				RB_R	GCG ATA TAA TCA CCG CCG TCG		
AE1	Anion Exchanger Like 1	39943	Bicarbonate transporter, SLC4 family	AE1_F	TTC ACG CTC TTC CAG TTC TC	102	Madkinder et al. 2011
				AE1_R	GAG GAA GGC GAT GAA GAA TG		
αCA	α Carbonic Anhydrase 2	456048	Alpha carbonic anhydrase	αCA2_F	AGA GCA GAG CCG TAT CAA CA	134	Richier et al. 2011
				αCA2_R	TCC TCT CGA AGA GCT GGA A		
δCA	δ Carbonic Anhydrase	436031	Delta carbonic anhydrase	δCA_F	ACC AGC ACC AGA TST TCA AG	87	Bach et al. 2013
				δCA_R	TCT CCG CAA CCA TCA TCT C		
CAX3	Ca <sup>2+</sup> /H <sup>+</sup> exchanger 3	416800	Ca <sup>2+</sup> /H <sup>+</sup> exchangers, similar to CAX family	CAX3_F2	CTC CTC TGC GTC TTT GCA T	90	Madkinder et al. 2011
				CAX3_R2	GAG GGC GGT GAT GAG GTA		
ATPVc/c	Vacuolar-type H <sup>+</sup> pump	359783	Vacuolar H <sup>+</sup> -ATPase, VD, subunit c/c'	ATPV_F	TAC GGC ACT GCA AAG TCT G	88	Madkinder et al. 2011
				ATPV_R	ACC GGG ATG ATG GAC TTC		
PATP	Plasma membrane type H <sup>+</sup> pump	67081	P type H <sup>+</sup> -ATPase	PATP_F	GAG CAC AAG TTC CTC ATC GTC	105	Bach et al. 2013
				PATP_R	CAG GTC GGC CTT GTT GAG		
NhaA2	Na <sup>+</sup> /H <sup>+</sup> exchanger 2	447659	Na <sup>+</sup> /H <sup>+</sup> antiporter	NhaA2_F	CTG GTG TGG TAT GGC ATC TC	80	Bach et al. 2013
				NhaA2_R	GTT GCT GGC GTC CAT TC		
LCIX	Low CO <sub>2</sub> induced gene	457739	Protein in <i>Emiliania huxleyi</i> 457739	LCIX_F	CAG CAG TCG TGG CTC AAG	94	Bach et al. 2013
				LCIX_R	CGT AAG CGA CGT GGA TCA G		
GPA	Ca <sup>2+</sup> binding protein	431830	Calcium-binding protein in <i>Emiliania huxleyi</i>	GpABR_F	AGG CCT TCT CCA GCA TCA T	70	Richier et al. 2009
				GpABR_R	GTT CAG GGT GCT CTC CGA G		



# Generic measuring issues

- Considerably **different units** across the different techniques
- Measurements tend to **need to be normalised**
  - organism: surface area, skeletal weight, body mass, biomass...
  - communities: volumetric, surface area...
- **Not trivial to compare!**
- Most measure **NET** calcification – difficult to disentangle the impacts on the organisms ability to calcify with dissolution.



# Summary

- Calcification ability has a **connection to energy** budgets
- **Feeding rates** may overcome some of the costs –will food supply change?
- **Dissolution rates** will increase as saturation state decreases – important for exposed material
- **Bio-erosion** may also further impact of OA
- **Adaptation** potential?
- **Interactions** between organisms
- Complexity of **multiple stressors**